

## **Protocol 501\_Mouse Albumin ELISA**

### **Reagents**

- ELISA coating buffer: 0.1M NaHCO<sub>3</sub> (3.18g Na<sub>2</sub>CO<sub>3</sub> + 5.88g NaHCO<sub>3</sub> made up to 1000ml with water).
- ELISA blocking buffer (PBS, 2% BSA – NB NO AZIDE – really inhibits HRP!!!!)
- ELISA wash buffer (PBS, 1:2000 Tween-20)
- ELISA substrate buffer: 0.1M NaH<sub>2</sub>PO<sub>4</sub>, pH 4 (13.8g NaH<sub>2</sub>PO<sub>4</sub> made up to 1000ml)
- Polyclonal Goat-anti-mouse Albumin (Bethyl Labs Cat# A90-134A)
- Polyclonal Goat-anti-mouse Albumin HRP (Bethyl Labs Cat# A90-134P)
- Standard – Mouse reference serum (Bethyl Labs Cat# RS10-101) Contains 45mg/ml albumin. Use top concentration of 1µg per ml i.e. dilute 1:45000! 1:450 per-diluted stock can be made, aliquoted and stored at -20 until needed for 1:100 dilution is this is easier)
- Nunc immunoplates

### **Method – as for isotype ELISA but:**

COAT plates in normal ELISA-coating buffer at pH 9.6 with 1:200 of unlabeled goat-anti-mouse albumin

Block with 2% BSA (no problem with cross-reactivity) in PBS

Add in samples starting at around 25mg/ml of cecal content and dilute 3x down the plates in duplicate

Add standards starting at 1µg/ml (reference serum contains 45mg/ml therefore dilute 1:45000)

Add developing antibody 1:33000 in PBS/BSA

### **Method – isotype ELISA**

1. Coat plates:
  - a. Dilute coating antibody 1:1000 in ELISA coating buffer
  - b. Plate 50µl per well. Tap the plate to ensure even spreading.
  - c. Cover the plates in parafilm and incubate 6hrs (rm temp) to overnight (4°C) in a humidified box (wet tissue at the bottom!)
2. Block:
  - a. Wash plates 3 times by immersion in ELISA wash buffer
  - b. Add 150µl of 2% BSA/PBS per well. Incubate for 15mins to several hours (plates can be frozen like this and stored long-term)
3. Add samples:
  - a. Plan out how to plate your samples and standards. (page 3!)
  - b. Make standard dilutions of standards and samples in blocking buffer, leaving a final volume in the wells of (60µl x the number of different ELISAs to be run on each sample).

- c. Flick out blocking buffer and pat plates dry
  - d. Pipette 50 $\mu$ l of sample/standard per well
  - e. Incubate for 3hrs (rm temp) to overnight (4°C) in a humidified chamber, sealed with parafilm
4. Develop plates
- a. Wash 6x by immersion in ELISA wash and pat dry
  - b. Dilute HRP-anti-isotypes 1:1000 in blocking buffer
  - c. Add 50 $\mu$ l per well and incubate for 1hr at rm temp
  
  - d. Wash 6x by immersion in ELISA wash and pat dry.
  - e. Make up 10ml substrate per plate (10ml substrate buffer, 1mg ABTS, 5 $\mu$ l H<sub>2</sub>O<sub>2</sub> – do not allow this to sit for more than about 10mins after the H<sub>2</sub>O<sub>2</sub> has been added)
  - f. Add 100 $\mu$ l substrate per well
  - g. Incubate for at least 15mins then read A405 on ELISA reader