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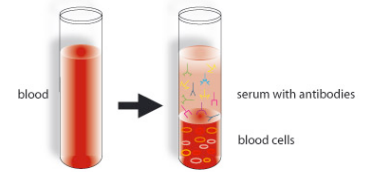
## Protocol 509\_IgE ELISA mouse serum

### Sample collection:

Collect blood into serum tube (brown cap)

Centrifuge serum for 10' at 10.000 g to separate cells, fibrinogen and other proteins involved in coagulation from serum (supernatant)

Store sample at -20°C until performing ELISA



### Method:

#### Day 0

**COATING** (up to 7 samples per 96 well plate)

1. Dilute **anti-IgE Ab (6HD5)** in ELISA coating buffer to 5 µg/ml  
Calculate 5 ml coating buffer/plate
2. Add 50 µl/well to ELISA plates (96 well, flat bottom,)
3. Wrap in saran, O/N at 4°C

#### Day 1

1. Flick off unbound coating antibody
2. Wash plate 3× with ELISA washing buffer

#### BLOCKING

3. Block plates by adding 100 µl/well 5% BSA, PBS
4. Incubate **2 h at RT**

#### LOADING SAMPLES

5. Flick off blocking solution, no need to wash here
6. Titrate samples on plates (see plate layout below)
  - a. Put 110µl 2.5 % BSA in PBS into A2-A12
  - b. Put 55 µl 2.5 % BSA in PBS into B1-H12
  - c. Put 165 µl **standard IgE at 5000 ng/ml** diluted in 2.5 % BSA in PBS into A1, titrate 55µl to A12 (1:3 titration)  
*for standard in row A titrate to column 12*
  - d. Put 55 µl serum samples 1-7 into B1-H1
  - e. Titrate for all rows (B-H) 55 µl across the plate to column 11, discard last 55 µl (serum 1:2 titration).
7. Incubate **2h at RT**
8. Wash plates 3× with ELISA washing buffer

#### SECONDARY ANTIBODY

9. Add **50 µl/well of secondary ab: anti-IgE(RIE4)-biotin** diluted in PBS 2.5% BSA to 2.5 µg/ml
10. Incubate **1.5h at RT** (or O/N at 4°C)
11. Wash plates 3× with ELISA washing buffer

#### STREPTAVIDIN

12. Add **100 µl/well streptavidin-HRPO** -Pat no 890803 R&D systems- (horseradish peroxidase) at 1 µg/ml (1:500 in PBS 2.5% BSA)
13. Incubate **max 1 h at RT** (35 min to max 1 h)
14. Wash plates 3× with ELISA washing buffer

#### DEVELOPING PLATE

15. Add **100 µl/well Reaction buffer**





### PLATE READING and analysis with MPM III 1.8

- ELISA reader is on G floor (Switch on, password 00000)
- Open MPM III software on computer next to ELISA reader
- File>New experiment>Read plate
- Reading mode: Endpoint; Mix time(s): 3: mixing speed: L
- click “start read”
- after plate has been read check whether standard curve is S-shaped:
- file>open protocol>IgE protocol
- then go to “curve fit” and check curve
  - if it is still linear, wait a few minutes longer and read again
  - if the curve fit is good: r2 0.9-1, save file!

### To analyse:

- Open data file and IgE protocol
- go to: “Report”
- use value  $Y = \dots / 0.345 / \dots$  to get the value of your sample in the titration series that fits the linear standard curve and therefore reflects the true IgE level best
- the IgE concentration is given in **ng/mL**

### File:

IgE protocol for the above plate layout is named: IgE protocol and saved as 510\_IgE protocol

Number of plates	Reagent	stock conc		end conc		Vf (ml)	Vi (µl) ?
1	<b>Coating antibody 6HD5</b>	5,43	mg/ml	5	µg/ml	5	4,60405157
2		5,43	mg/ml	5	µg/ml	10	9,20810313
3		5,43	mg/ml	5	µg/ml	15	13,8121547
4		5,43	mg/ml	5	µg/ml	20	18,4162063
5		5,43	mg/ml	5	µg/ml	25	23,0202578
6		5,43	mg/ml	5	µg/ml	30	27,6243094
1	<b>Secondary antibody RIE-4</b>	1,3	mg/ml	2,5	µg/ml	5	9,61538462
2		1,3	mg/ml	2,5	µg/ml	10	19,2307692
3		1,3	mg/ml	2,5	µg/ml	15	28,8461538
4		1,3	mg/ml	2,5	µg/ml	20	38,4615385
5		1,3	mg/ml	2,5	µg/ml	25	48,0769231
6		1,3	mg/ml	2,5	µg/ml	30	57,6923077

