Protocol for importation of materials after setting up an isolator

To bring an isolator into use it must first be sterilized as detailed on the Isolator setup protocol. Once this has been done, materials need to be imported into the isolators after first autoclaving them in the transport rings. To simplify this process we have drawn up the following list of materials to import into the isolator in sequence.

Stages of the preparation

1. Initial isolator setup (see **Gnoto1000**)
2. First transport cylinder connection including taking samples to confirm sterility (see detailed instructions below)

Materials for first transport cylinder connection

A. Inside the drum

1. 10 Cages
2. 10 Hoppers
3. 10 Water bottles & sipper caps
4. Dustpan and brush
5. 10 Paper bags
6. 5 J cloths for cleaning (cut in half)
7. Small roll of masking tape
8. 1 pair of scissors (no points)
9. 1 large pair of forceps
10. 6x 100ml Duran flasks contained within bottle holder each marked with serial number[1]
11. 9x Gauze swabs
12. Sponge filter and wire underlay for the exhaust port.
13. 134°C chemical indicator strip
14. Mouse tail holding forceps
15. SPF cage for food
16. Nestlets
17. Wide funnel to assist loading
18. Metal I.D. tags and hooks

B. Additional materials required

1. Transport drum and second to bottom tray as base (use drums 3 or 4 with no shelves, larger drum)
2. Mylar film
3. Vinyl tape[2]
4. Free autoclave time (make reservation ahead of time)
5. Spray gun and hose

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[1] Coding for the sample bottles. These are in groups of 6 and will include the year, month, date, and serial number, **coded in the standard fashion** YYYYMMDD# where YYYY = the 4 digit year, MM = the 2 digit month, and DD = the 2 digit date. The # symbol is replaced with the serial number and may be as many digits as necessary. All sample bottles will thus have at minimum a 9-digit code.

[2] **Colour Codes for Vinyl Tape**
Red – Cages & Equipment
Blue – Water
Green – Bedding & Food

6. Spray gun container with 100ml of 2% peracetic acid (freshly made up). The lid on the container should be tightly screwed down.
7. Swab loaded into swab holder
8. 250ml Duran flask to hold 10 ml of peracetic acid.
9. Form for cylinder connection (GnotoformABC.xls, where ABC is the three-digit isolator code[3]—see **Gnoto1010** for the codes)
10. Spill container (rat cage) with absorbent pad

Procedure for first transport cylinder connection

1. Clean out the transport drum and remove any old tape.
2. Put the second tray into the drum to provide a platform for the materials, or use drum 3 or 4 (these work best).
3. Load the drum with the materials in List A above.
4. Seal the drum with Mylar film (**see Gnoto1023**).
5. Autoclave the drum on appropriate program (**see Gnoto1011**).
6. Remove the drum from the autoclave and bring into the gnotobiotic unit (**see Gnoto1007**).
7. Connect the drum to the isolator as per **Gnoto1024**.
8. Import materials and arrange them neatly.
9. Put the filter and overlying sponge on the exhaust ventilation port.
10. Take samples for bacterial culture. Two bottles should remain unopened. The remaining four bottles should be opened and swabs for two of them wiped on the floor and sides of the isolator, and for the other two in the middle of the quadralock and the input and exhaust ventilation ports and all small-hole ports. After wiping these areas, the place the swabs in the bottles and seal by shutting the screw caps. The bottles are then transferred back to the transport cylinder.
11. Close inner quadralock door.
12. Detach the sleeve and place it to be cleaned prior to next use.
13. Complete the form GnotoformABC.xls, print off a copy and Email it to Jorum
14. Place the sample bottles in the tray in the corridor.
15. Store the transport cylinder.
16. Repeat steps 1-15 for subsequent transport cylinder connections.

[3] For example, Gnotoform101.xls refers to isolator 101, Gnotoform102.xls refers to isolator 102, and so on.